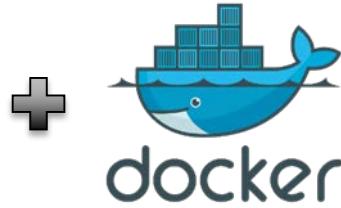


Doceexpress –

A galaxy docker for estimation of Expression profiling in RNA-seq



SPEAKER: Ping-Heng Hsieh
DATE: 2018/12/07



- » Introduction for Docexpress
- » Walkthrough – Estimate Expression Profiling
- » Introduction for Docmethyl

1.

INTRODUCTION FOR DOCEXPRESS

Introduction for Docexpress

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Docexpress

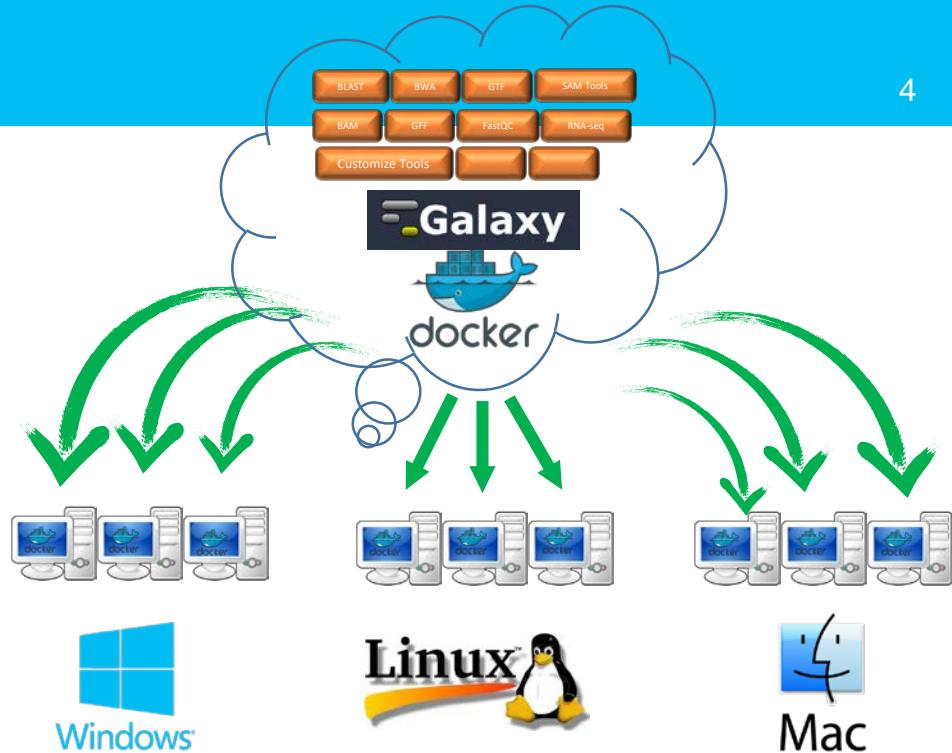
- » Galaxy + Docker

Galaxy

- » “Galaxy is an open source, web-based platform for data intensive biomedical research.” – by Galaxy

Docker

- » “Docker is an open platform for developing, shipping, and running applications.” – by Docker docs



Docexpress

- » For alleviating the burden on processing massive NGS data, we create the workflows based on galaxy/ Docker to estimate expression profiling in RNA-seq.

- » After the data pre-processing done, the expression profiling can be submitted to MOLAS, is a robust web application that can take gene expression data (FPKM/TPM) from different libraries as inputs, map these expressed genes with build-in annotations for further analyses and reveal biological meaning of the complex data in the intuitive interface.



1. Install Docker on your local system [[Ref.](#)]
2. Open a terminal or Use command mode
3. Following the “Install & Usage” on our docker hub [[link](#)]

- » Create data store directory “galaxy_guest”

md galaxy_guest #for windows command line

- » Pull the docexpress images

docker pull lsbnb/docexpress_fastqc

- » Run docexpress

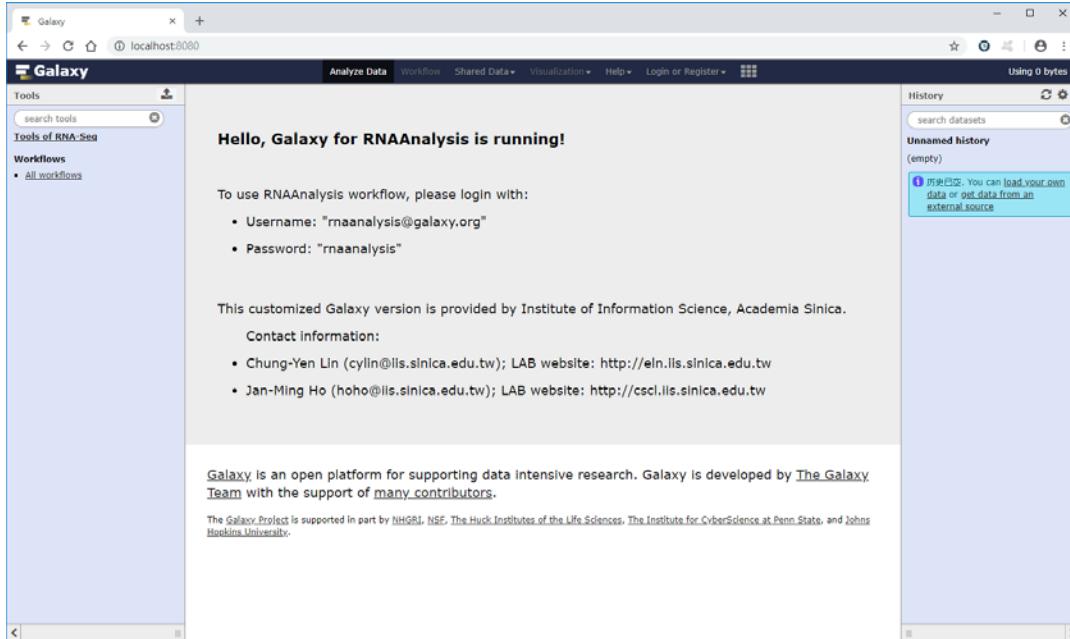
*docker run -d -t -i -p 8080:80 -p 8021:21 -p 8022:22 -v
%cd%/galaxy_guest:/root/galaxy/database/ftp/rnaanalysis@galaxy.org/
lsbnb/docexpress_fastqc /bin/bash*

- » Open the browser and input the address

“http://localhost:8080/”

Lunch succeeded!

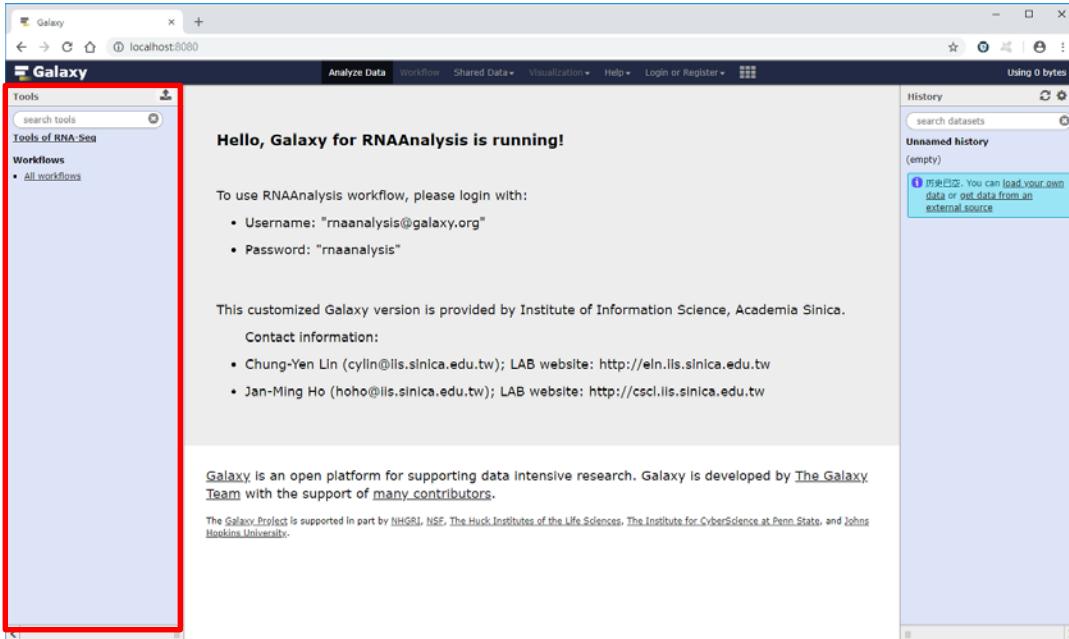
8



Lunch succeeded!

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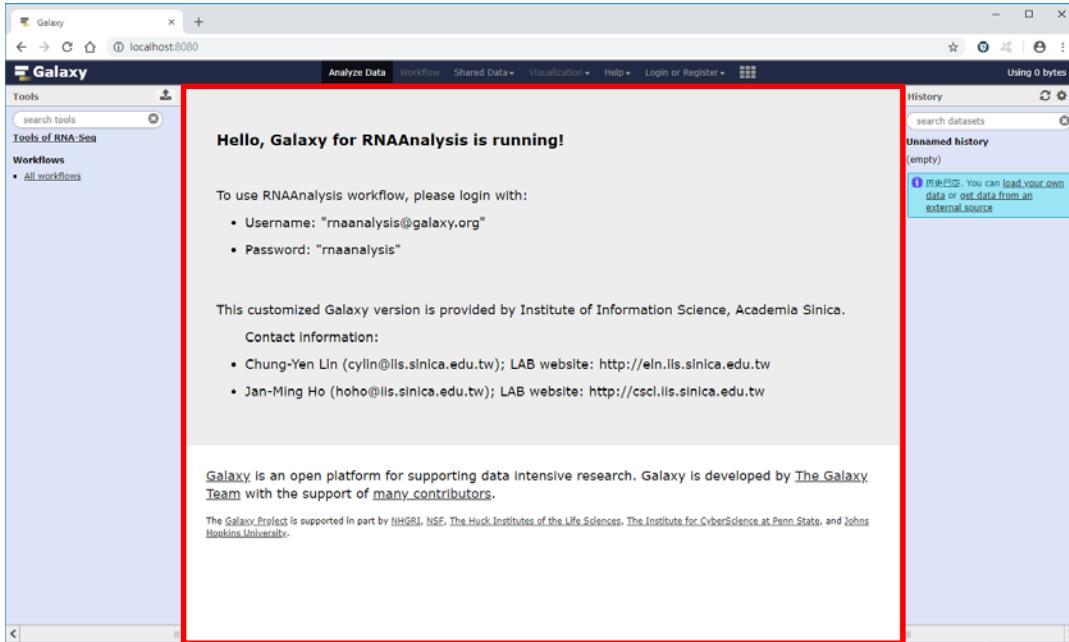
Tool panel: which contains all tools and workflows



Lunch succeeded!

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Main : show the homepage and execution result...etc

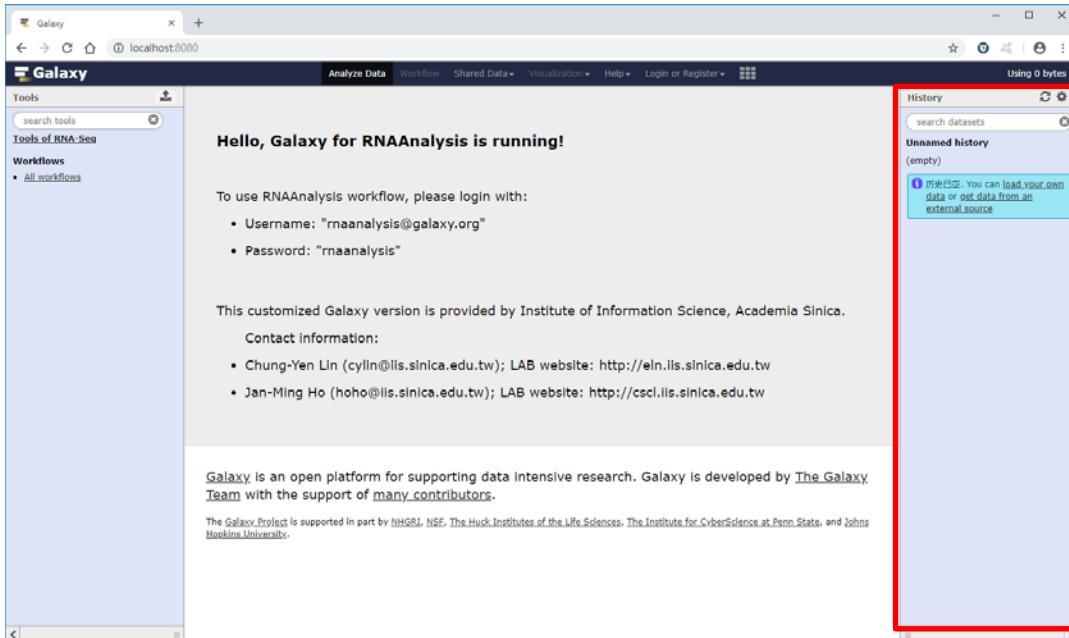


Lunch succeeded!

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History: contains all input and output data and result file.

Please get on Galaxy 101 for more details [[link](#)]



2.

WALKTHROUGH

– Estimate Expression Profiling

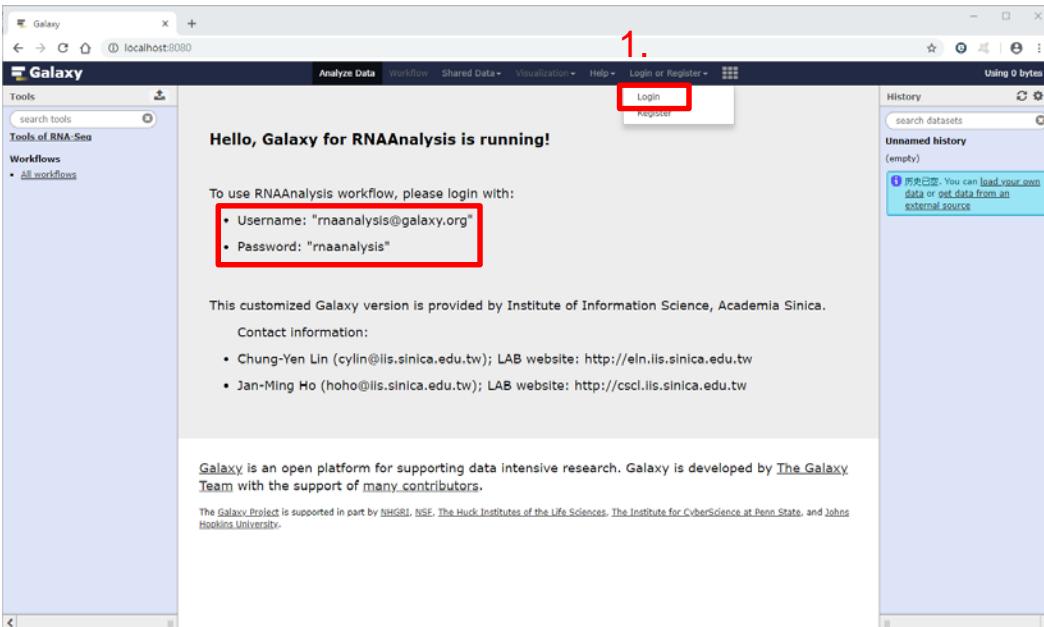
Download test data

- » Download [link]
- » Put all test data into directory “galaxy_guest”

Login Galaxy

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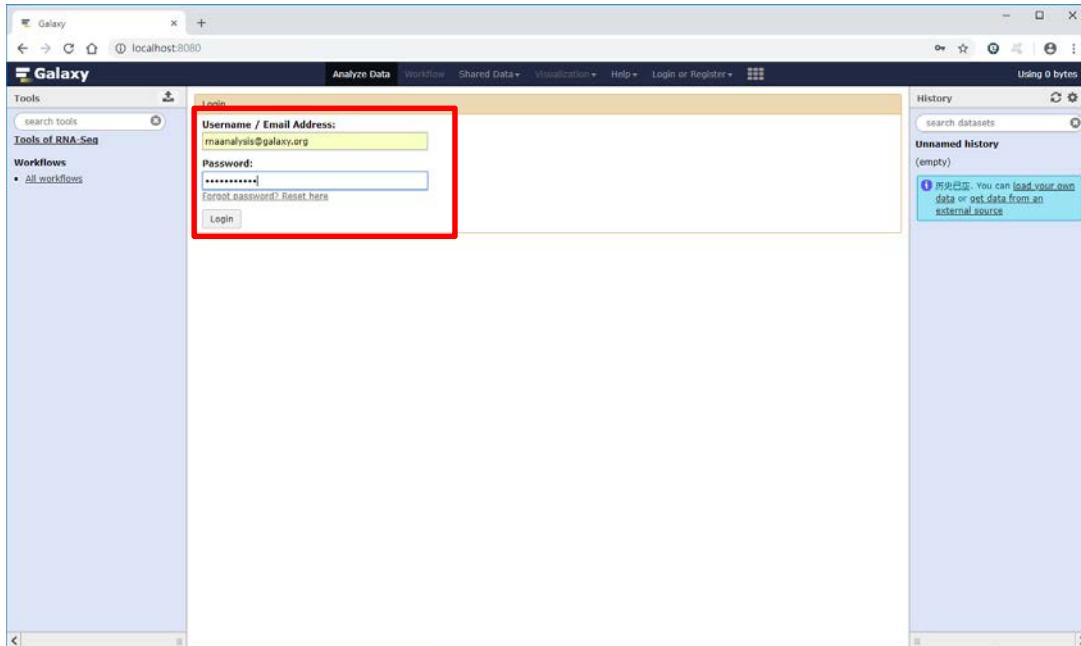
Login for full accessing all functions of Docexpress



Login Galaxy

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Login for full accessing all functions of Docexpress



Upload files

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Two ways for calling the upload function

The screenshot shows the Galaxy web interface running at localhost:8080. The main content area displays a message: "Hello, Galaxy for RNAAnalysis is running!" and instructions for logging in with a username and password. Below this, contact information for the Institute of Information Science, Academia Sinica, is provided. At the bottom, there is a note about the Galaxy project and its contributors.

Tools (highlighted with a red box)

- search tools
- Tools of RNA-Seq
 - FastQC: Read Quality reports
 - Trim Galore!: Quality and adapter trimmer of reads
 - Column Join on Collections
 - StringTie merge transcripts
 - Upload file from your computer** (highlighted with a red box)
 - Sort data in ascending or descending order
 - Cut columns from a table
 - StringTie transcript assembly and quantification
 - HISAT2: A fast and sensitive alignment program
- Workflows
 - All workflows

History

search datasets

Unnamed history (empty)

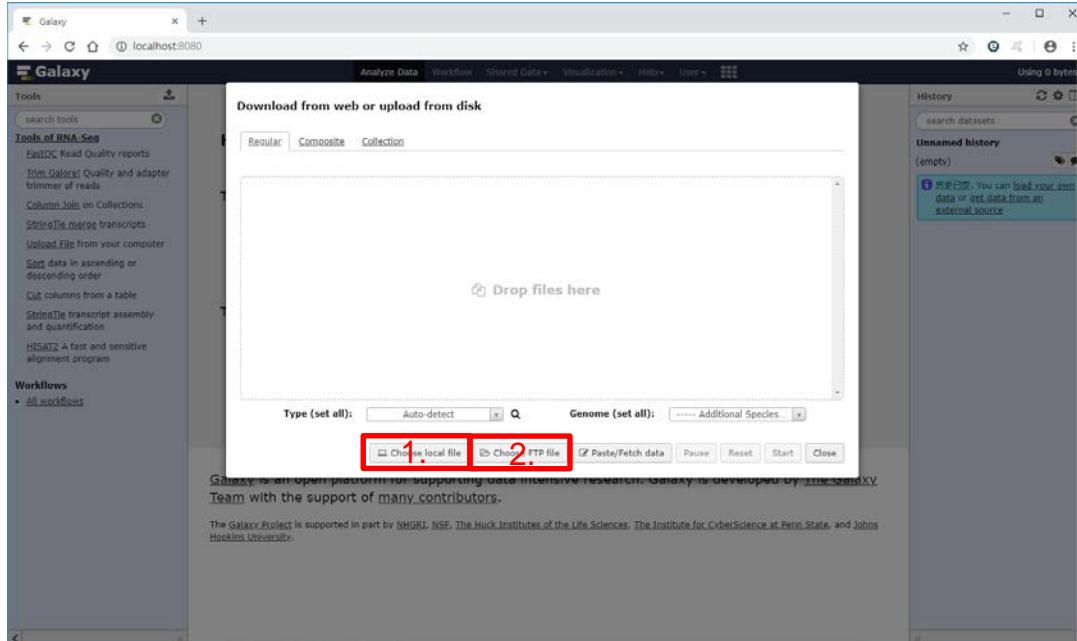
历史已空。您可以从外部源加载自己的数据或从外部源获取数据。

Upload files

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Two ways for uploading data,

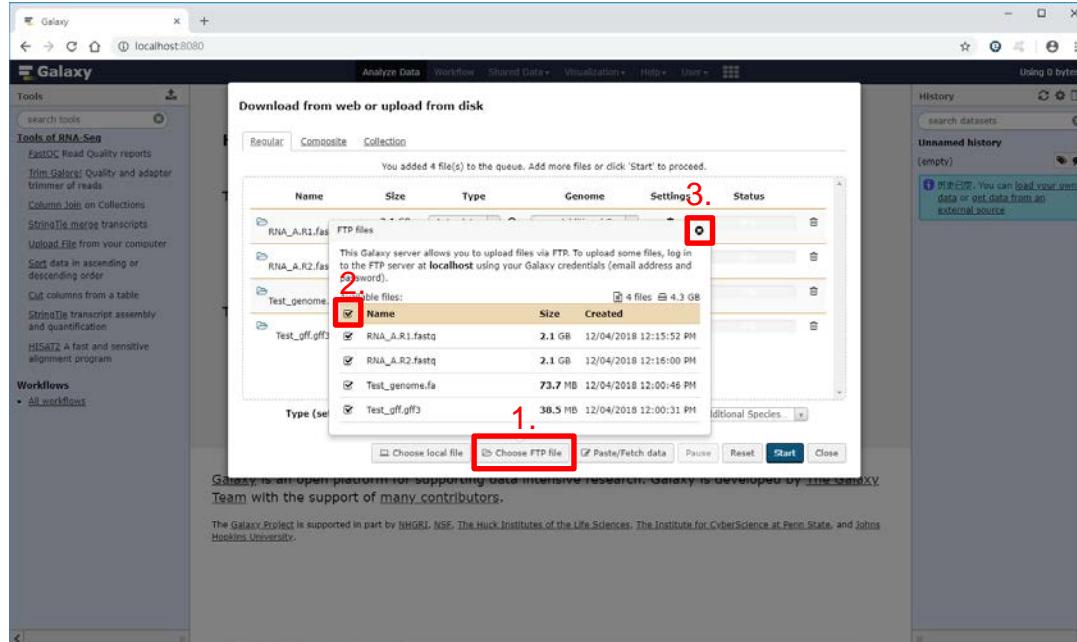
1. Single file < 2 GB
2. No limitation, depend on the capability of local system



Upload files

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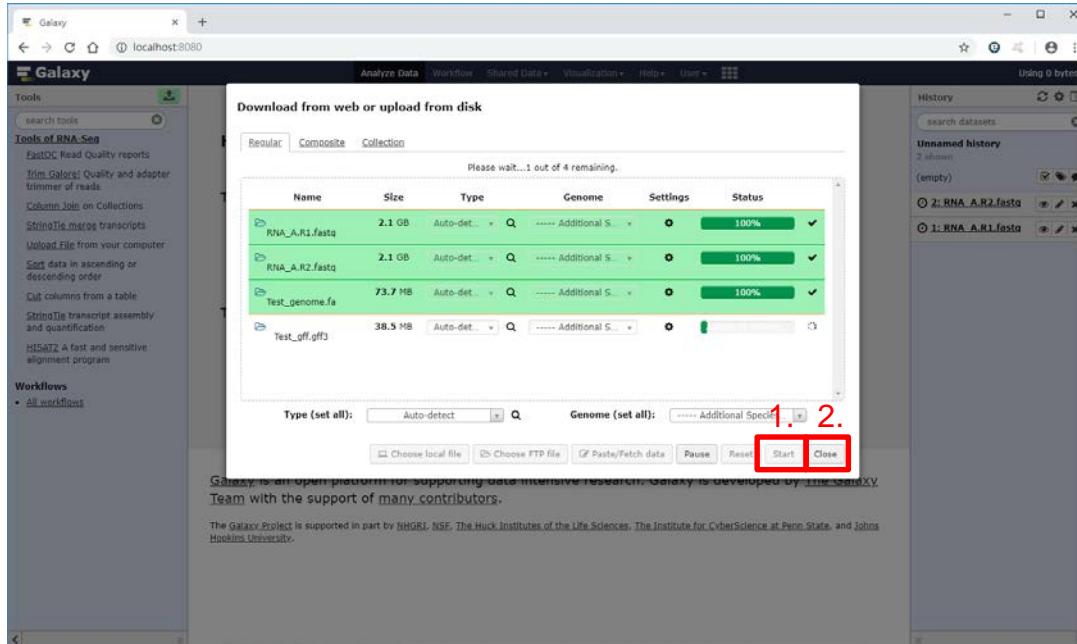
1. Choose FTP file
2. Check all files
3. Close selection window



Upload files

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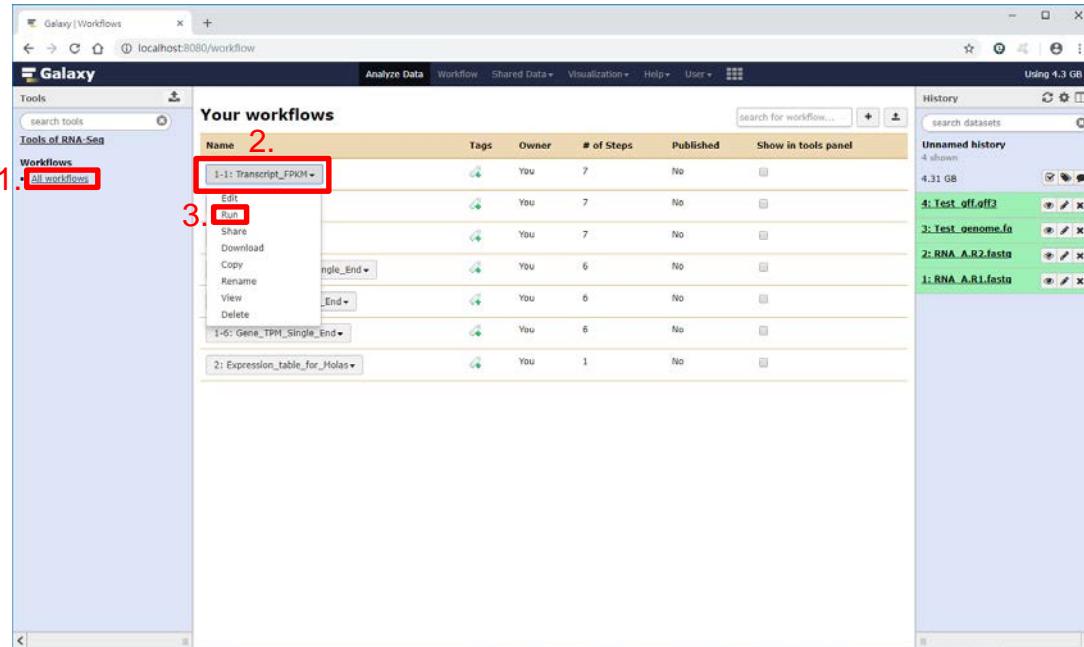
1. Upload files
2. Close upload function



Run Workflow : 1-1: Transcript_FPKM

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1. Choose “All workflows”
2. Choose “1-1: Transcription_FPKM”
3. Run the workflow



Run Workflow : 1-1: Transcript_FPKM

21

1. Choose RAN-seq data as the input

Workflow: 1-1: Transcript_FPKM

Run workflow

History Options
Send results to a new history
 Yes No

Trim Galore! (Galaxy Version 0.4.3.1)

Is this library paired- or single-end?
Paired-end

1. Reads in FASTQ format
1: RNA_A.R1.fastq

2. Reads in FASTQ format
2: RNA_A.R2.fastq

Adapter sequence to be trimmed
Automatic detection

Trims 1 bp off every read from its 3' end.
false

Remove N bp from the 3' end of read 1
Empty.

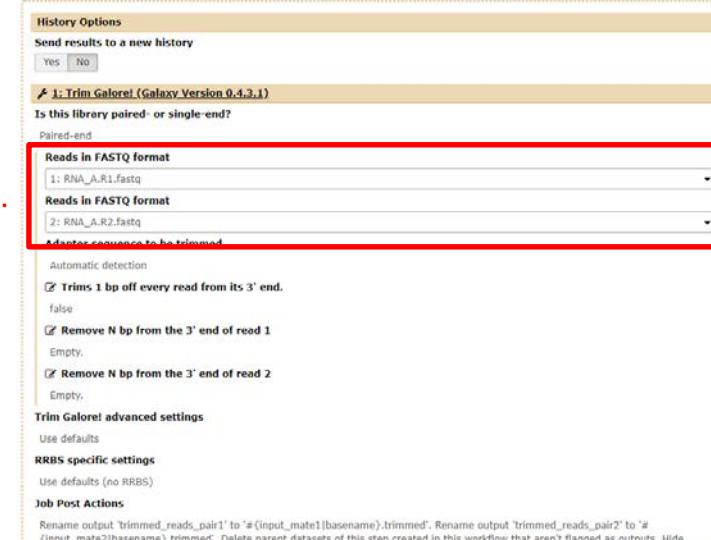
Remove N bp from the 3' end of read 2
Empty.

Trim Galore! advanced settings
Use defaults

RRBS specific settings
Use defaults (no RRBS)

Job Post Actions
Rename output 'trimmed_reads_pair1' to '#{input_mate1}trimmed'. Rename output 'trimmed_reads_pair2' to '#{input_mate2}trimmed'. Delete parent datasets of this step created in this workflow that aren't flagged as outputs. Hide

1.



Run Workflow : 1-1: Transcript_FPKM

22

1. Choose RAN-seq data as the input
2. Select genome

Workflow: 1-1: Transcript_FPKM

Run workflow

2: HISAT2 (Galaxy Version 2.1.0)

Source for the reference genome

Select the reference genome

3: Test_genome.fa

Single-end or paired-end reads?

Paired-end

FASTA/Q file #1

Output dataset 'trimmed_reads_p1' from step 1

FASTA/Q file #2

Output dataset 'trimmed_reads_p2' from step 1

Specify strand information

Forward (FR)

Paired-end options

Use default values

Summary Options

Advanced Options

Job Post Actions

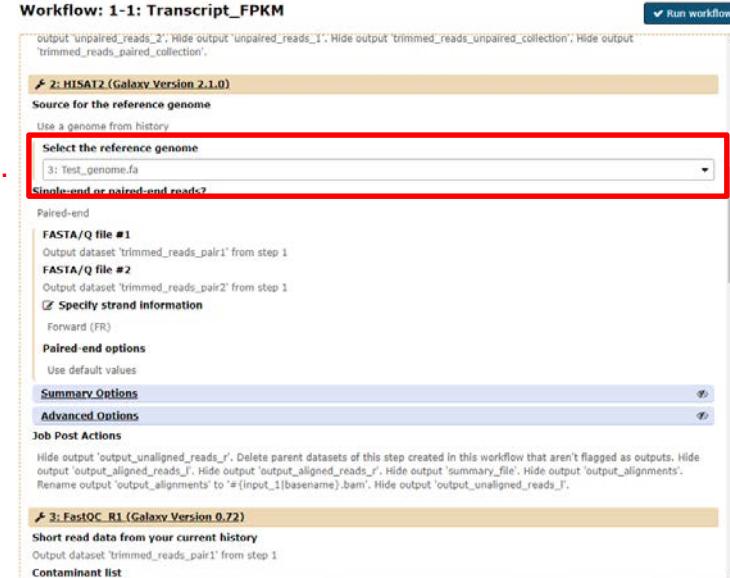
Hide output 'output_unaligned_reads_r'. Delete parent datasets of this step created in this workflow that aren't flagged as outputs. Hide output 'output_aligned_reads_l'. Hide output 'output_aligned_reads_r'. Hide output 'summary_file'. Hide output 'output_alignments'. Rename output 'output_alignments' to '#(input_1 basename).bam'. Hide output 'output_unaligned_reads_l'.

3: FastQC_R1 (Galaxy Version 0.72)

Short read data from your current history

Output dataset 'trimmed_reads_p1' from step 1

Contaminant list



Run Workflow : 1-1: Transcript_FPKM

23

1. Choose RAN-seq data as the input
2. Select genome
3. Select GFF/GTF file
4. Run workflow

Workflow: 1-1: Transcript_FPKM

4. Run workflow

5: StringTie (Galaxy Version 1.3.3.1)

Input mapped reads
Output dataset 'output_alignments' from step 2

Specify strand information
Unstranded
Select 'Forward (FR)' if your reads are from a forward-stranded library, 'Reverse (RF)' if your reads are from a reverse-stranded library, or 'Unstranded' if your reads are not from a stranded library. See Help section below for more information. Default: Unstranded

Use a reference file to guide assembly?

Use reference GTF/GFF3

Reference file
Use a file from history
3. GTF/GFF3 dataset to guide assembly
4: Test_gff.gff3
 Use Reference transcripts only

true

Output files for differential expression?
Ballgown
 Output coverage file?
false

Advanced Options

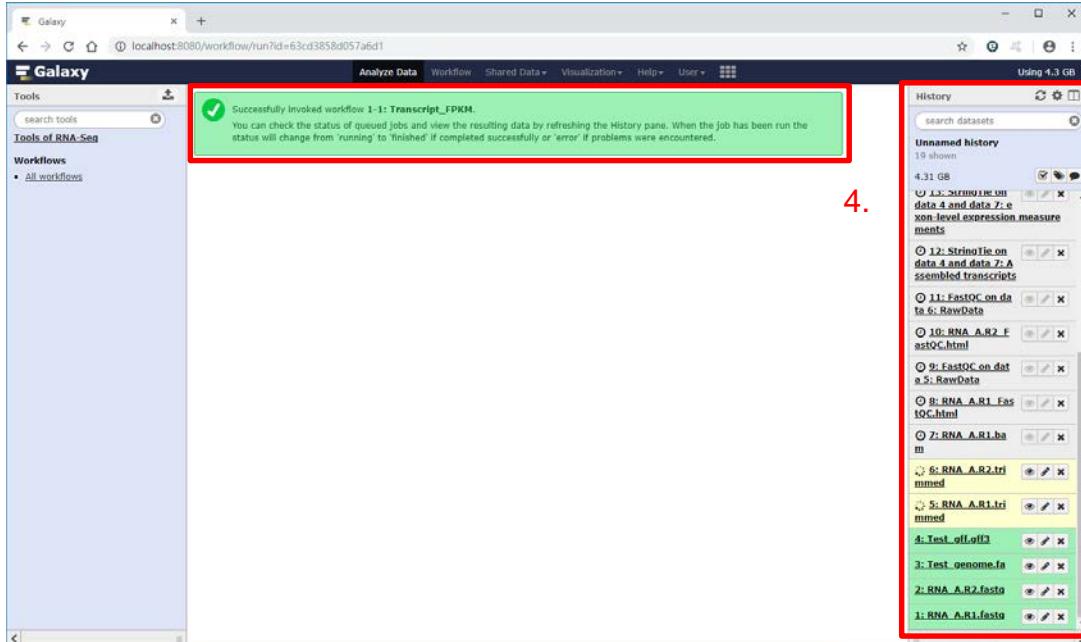
Job Post Actions
Hide output 'gene_counts'. Hide output 'intron_transcript_mapping'. Delete parent datasets of this step created in this workflow that aren't flagged as outputs. Hide output 'coverage'. Hide output 'output_gtf'. Rename output 'transcript_expression' to 'a' ('input_bam basename'). Hide output 'transcript_counts'. Hide output 'legend'. Hide output 'gene_abundance_estimation'. Hide output 'exon_expression'. Hide output 'transcript_expression'. Hide output 'exon_transcript_mapping'. Hide output 'intron_expression'.

6: Sort (Galaxy Version 1.0.3)

Run Workflow : 1-1: Transcript_FPKM

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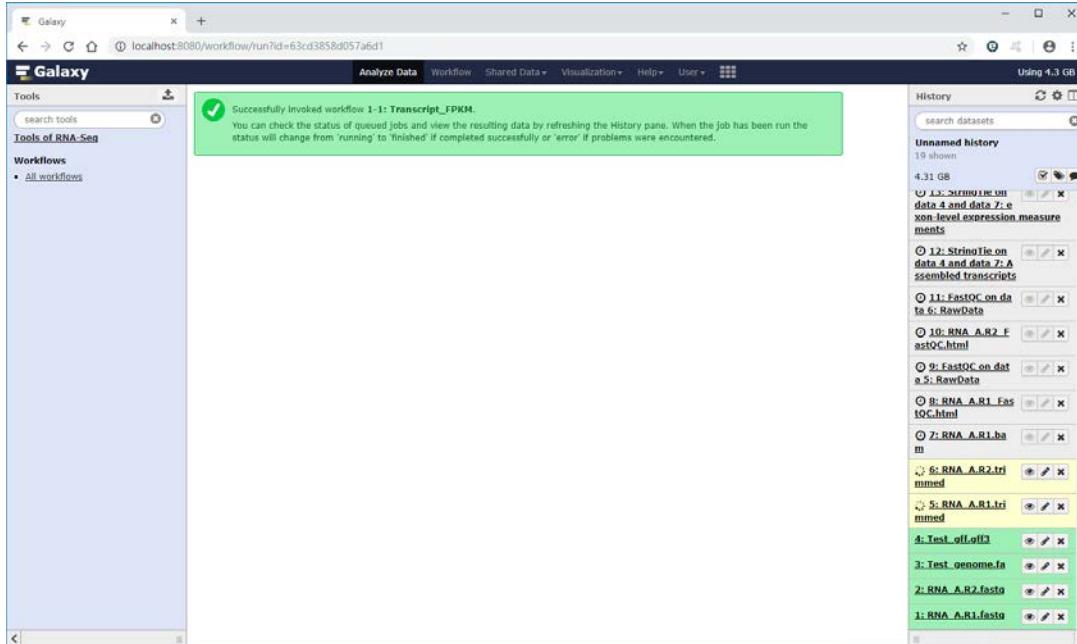
1. Choose RAN-seq data as the input
2. Select genome
3. Select GFF/GTF file
4. Run workflow



Run Workflow : 1-1: Transcript_FPKM

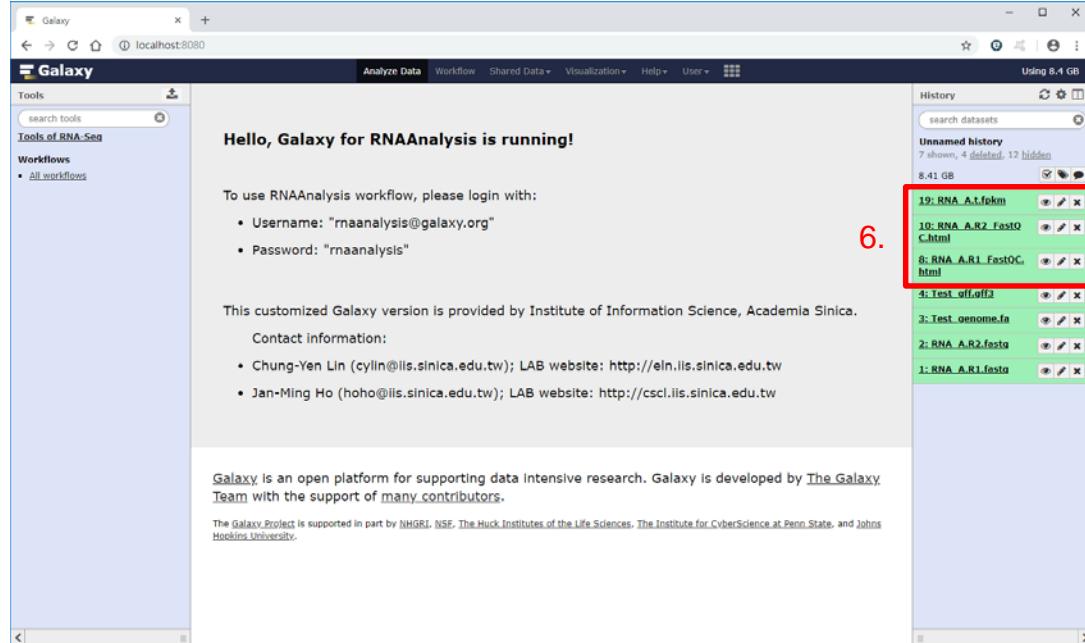
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1. Choose RAN-seq data as the input
2. Select genome
3. Select GFF/GTF file
4. Run workflow
5. Repeat 1. - 4. until all RNA-seq data selected and execute the workflow



Run Workflow : 1-1: Transcript_FFKM

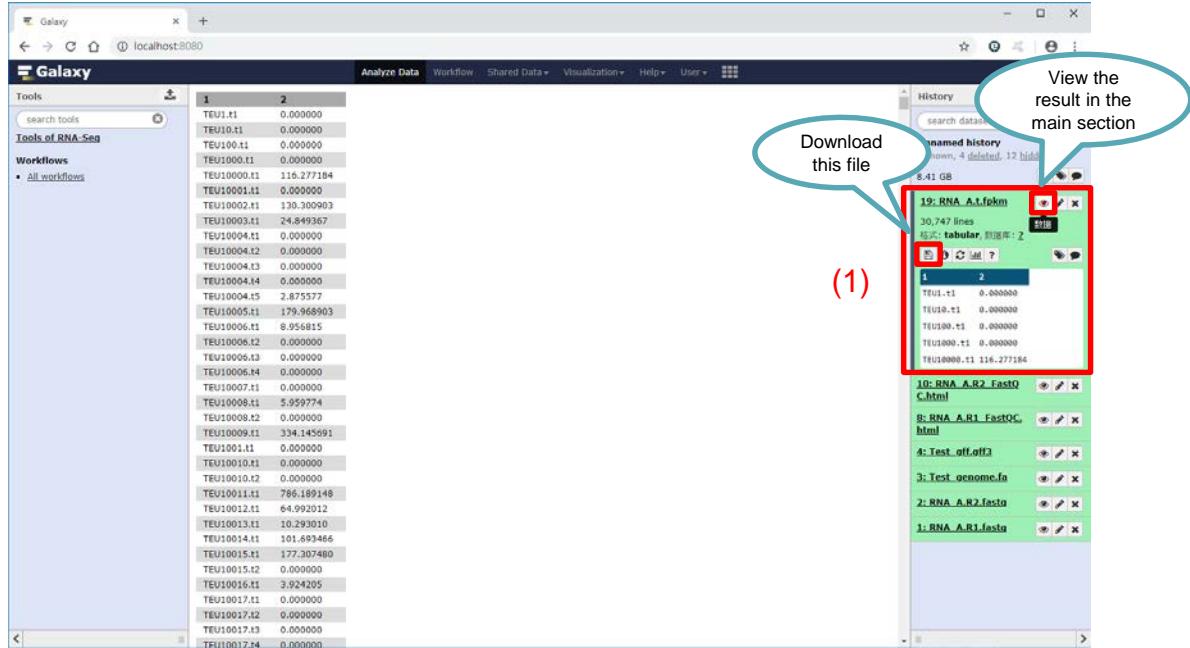
1. Choose RAN-seq data as the input
2. Select genome
3. Select GFF/GTF file
4. Run workflow
5. Repeat 1. - 4. until all RNA-seq data selected and execute the workflow
6. Outputs of workflow,
(1) FastQC.html
(2) Samlpe.t.fpkm



Output of Workflow : 1-1: Transcript_FFKM

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- Outputs of workflow,
(1) Samlpe.t.fpkm
(2) FastQC.html



Output of Workflow : 1-1: Transcript_FFKM

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- Outputs of workflow,
(1) Samlpe.t.fpkm
(2) FastQC.html

View the result in the main section

(2)

RNA_A_R1_trimmed FastQC Report

FastQC Report

Wed 5 Dec 2018

RNA_A_R1_trimmed

Summary

- Basic Statistics
- Per base sequence quality
- Per tile sequence quality
- Per sequence quality scores
- Per base sequence content
- Per sequence GC content
- Per base N content
- Sequence Length Distribution
- Sequence Duplication Levels
- Overrepresented sequences
- Adapter Content

Basic Statistics

Measure	Value
Filename	RNA_A_R1_trimmed
File type	Conventional base calls

History

- 1: RNA_A_R1.fastq
- 2: RNA_A_R2.FastQ.C.html
- 3: RNA_A_R1.html**
- 4: RNA_B_R2.fastq
- 5: RNA_B_R1.fastq
- 6: RNA_A_R2.fastq
- 7: RNA_A_R1.fastq

Run Workflow : 2: Expression_table_for_Molas

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1. Choose “All workflows”
2. Choose “2:Expression_table_for_Molas”
3. Run the workflow

The screenshot shows the Galaxy Workflow interface. On the left, there's a sidebar with 'Tools of RNA-Seq' and 'Workflows'. A red box labeled '1.' highlights the 'Edit workflows' button. In the center, the 'Your workflows' list is displayed. A red box labeled '2.' highlights the '2: Expression_table_for_Molas' workflow. A red box labeled '3.' highlights the 'Run' option in the context menu for this workflow. The right side of the interface shows a 'History' panel with several datasets listed.

Name	Tags	Owner	# of Steps	Published	Show in tools panel
1-1: Transcript_FPKM		You	7	No	
1-2: Gene_FPKM		You	7	No	
1-3: Gene TPM		You	7	No	
1-4: Transcript_FPKM_Single_End		You	6	No	
1-5: Gene_FPKM_Single_End		You	6	No	
1-6: Gene TPM_Single_End		You	6	No	
2: Expression_table_for_Molas		You	1	No	

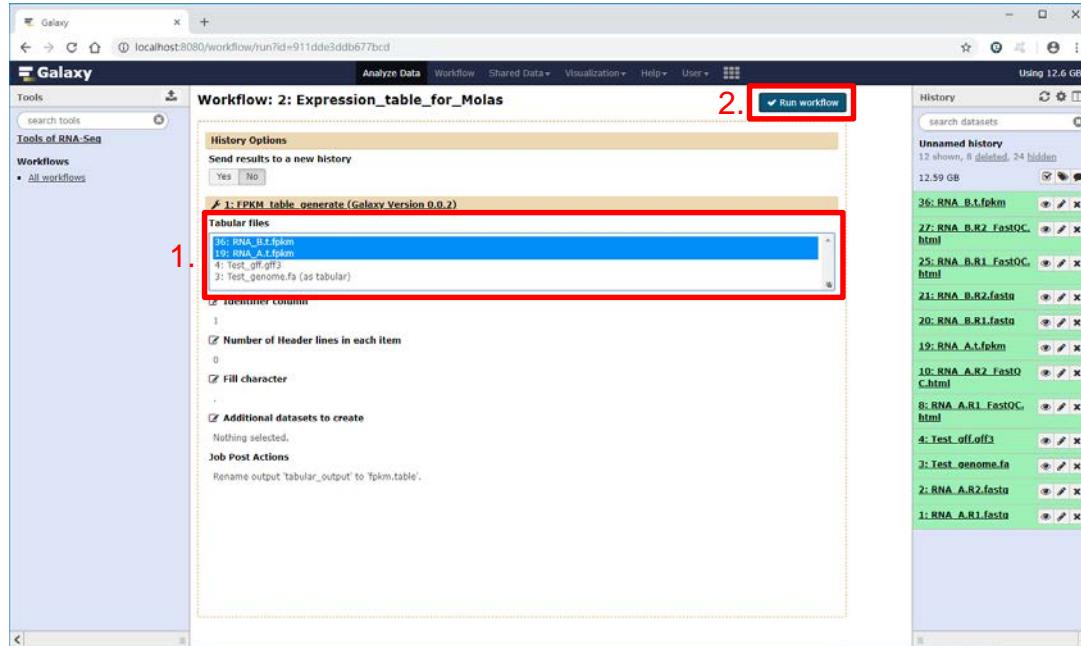
History:

- 36: RNA_B.tfpkm
- 27: RNA_B.R2_FastQC.html
- 25: RNA_B.R1_FastQC.html
- 21: RNA_B.R2.fasta
- 20: RNA_B.R1.fasta
- 19: RNA_A.tfpkm
- 10: RNA_A.R2_FastQC.html
- 8: RNA_A.R1_FastQC.html
- 4: Test.off.off1
- 3: Test_genome.fa
- 2: RNA_A.R2.fasta
- 1: RNA_A.R1.fasta

Run Workflow : 2: Expression_table_for_Molas

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1. Choose all .t.fpkm files with pressing the “Ctrl”
2. Run workflow



Run Workflow : 2: Expression_table_for_Molas

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1. Result: fpkm.table

The screenshot shows the Galaxy web interface at the URL `localhost:2080/workflow/run?id=911dde3ddb677bcd`. The main content area displays a green success message: "Successfully Invoked workflow 2: Expression_table_for_Molas. You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered." To the right, the "History" pane lists several datasets. A red box highlights the entry "37: fpkm.table", which is the result of the workflow. Other entries include "26: RNA_B.R1.fpkm", "27: RNA_B.R2_FastQC.html", "25: RNA_B.R1_FastQC.html", "21: RNA_B.R2.fasta", "20: RNA_B.R1.fasta", "19: RNA_A.t.fpkm", "10: RNA_A.R2_FastQC.html", "9: RNA_A.R1_FastQC.html", "4: Test.off.off3", "3: Test_genome.fa", "2: RNA_A.R2.fasta", and "1: RNA_A.R1.fasta".

Index	Dataset Name
37	fpkm.table
26	RNA_B.R1.fpkm
27	RNA_B.R2_FastQC.html
25	RNA_B.R1_FastQC.html
21	RNA_B.R2.fasta
20	RNA_B.R1.fasta
19	RNA_A.t.fpkm
10	RNA_A.R2_FastQC.html
9	RNA_A.R1_FastQC.html
4	Test.off.off3
3	Test_genome.fa
2	RNA_A.R2.fasta
1	RNA_A.R1.fasta

Run Workflow : 2: Expression_table_for_Molas

- View and download the fpkm.table for MOLAS system

The screenshot shows the Galaxy web interface at localhost:2080/workflow/run?id=911dde3ddb677bcd. The main area displays a table titled '#KEY' with columns 1, 2, and 3. The table contains numerous rows of data, mostly consisting of zeros. A red box highlights the 'fpkm.table' dataset in the history panel. To the right, a green preview window shows the first few rows of the table. Callouts indicate 'Download this file' pointing to the download icon in the preview window, and 'View the result in the main section' pointing to the main table area.

#KEY	RNA_A.t.fpkm_2	RNA_B.t.fpkm_2
TEU1.t1	0.000000	0.000000
TEU10.t1	0.000000	0.000000
TEU100.t1	0.000000	0.000000
TEU1000.t1	0.000000	0.000000
TEU10000.t1	116.277184	88.075256
TEU10001.t1	0.000000	0.000000
TEU10002.t1	130.309093	108.687370
TEU10003.t1	24.849367	0.000000
TEU10004.t1	0.000000	0.000000
TEU10004.t2	0.000000	1.210395
TEU10004.t3	0.000000	0.000000
TEU10004.t4	0.000000	0.000000
TEU10004.t5	2.675577	4.679134
TEU10005.t1	179.068903	953.210754
TEU10006.t1	8.956815	0.000000
TEU10006.t2	0.000000	0.000000
TEU10006.t3	0.000000	11.704576
TEU10006.t4	0.000000	0.000000
TEU10007.t1	0.000000	0.000000
TEU10008.t1	5.959774	0.000000
TEU10008.t2	0.000000	0.000000
TEU10009.t1	334.145491	194.510788
TEU10010.t1	0.000000	0.000000
TEU10010.t2	0.000000	0.000000
TEU10011.t1	786.189148	721.523882
TEU10012.t1	64.992012	99.557845
TEU10013.t1	10.293010	21.677700
TEU10014.t1	101.692466	147.884201
TEU10015.t1	177.307480	170.098160
TEU10015.t2	0.000000	0.000000
TEU10016.t1	3.924205	0.000000
TEU10017.t1	0.000000	0.000000
TEU10017.t2	0.000000	0.000000
TEU10017.t3	0.000000	0.000000

History:

- 37: fpkm.table
- 36: RNA_B.t.fpkm
- 27: RNA_B.R2_FastQ.html
- 25: RNA_B.R1_FastQ.html
- 21: RNA_B.R2.fasta
- 20: RNA_B.R1.fasta
- 19: RNA_A.t.fpkm
- 10: RNA_A.R2_FastQ.html
- 8: RNA_A.R1_FastQ.html

3.

INTRODUCTION FOR DOCMETHYL

DocMethyl

- » We packed a Docker container DocMethyl to deal with raw data processing, mapping, and methylation calling/ scoring to give the summary, mtable, of the whole genome methylation status by the gene.
- » Mtables are uploaded to the web server EpiMOLAS_web for linking with gene annotation databases that enable rapid data retrieval and analyses.



1. Install Docker on your local system [[Ref.](#)]
2. Open a terminal or Use command mode
3. Following the “Install & Usage” on our docker hub [[link](#)]

THANKS!

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Any questions?



1. Docker - <https://www.docker.com/>
2. Docker Install - <https://docs.docker.com/install/linux/docker-ce/ubuntu/>
3. Galaxy - <https://usegalaxy.org/>
4. Docexpress - <https://hub.docker.com/r/lisbnb/docexpress/>
5. DocMethyl - <https://hub.docker.com/r/lisbnb/docmethyl/>
6. Galaxy basic usage <https://galaxyproject.github.io/training-material/topics/introduction/tutorials/galaxy-intro-101/tutorial.html>